



A preliminary study on the occurrence of microscopic asexual fungi associated with bird nests in Brazilian semi-arid

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Abstract

Bird nests are made up of twigs, leaves and bark, among other things, all substrates that are decomposed by fungi. Thus, the richness of microscopic asexual fungi associated with the plant substrates that make up the bird nests in a fragment of the semi-deciduous seasonal forest in the “Serra da Maravilha”, Senhor do Bonfim, Bahia, Brazil was studied. The plant substrates were collected between February to July 2014. Forty-four species of microscopic asexual fungi were identified distributed in 36 genera, associated with the decomposition of 11 birds' nests. Among the species cataloged, *Actinocladium rhodosporium*, *Alternaria alternata*, *Cryptophiale kakombensis*, *Ellisembia adscendens*, *Gyrothrix circinata*, *Gyrothrix microsperma*, *Tetraploa aristata*, *Thozetella cristata* and *Torula herbarum* were found throughout the study, while the other species were found sporadically. *Dendryphion cubense* constitutes a new record for South America, *Endophragmiella valdiviana* represents a new record for Brazil and *Uberispora heteroseptata* is a new record for Bahia.

Keywords – biodiversity – birds – conidial fungi – mycota

Introduction

Fungi form an extremely abundant and morphologically diverse kingdom, which includes saprobic, symbiotic and parasitic species (Heredia et al. 2000). Microscopic asexual fungi are pioneers in decomposing plant materials since they have an enzymatic framework capable of breaking down the distinct substances found in these substrates (Dix & Webster 1995).

Estimates of the number of fungi on the planet, range up to 1.5 to 5.1 million species (Hawksworth 1991, Blackwell 2011), and there are many little or never explored environments. These studies suggest a great diversity of yet unknown fungi species, which are most likely in the tropical ecosystems. Thus, little explored micro-habitats like bird nests make an ideal environment for fungal colonization since they have a diversified plant composition.

This work aimed to estimate the richness of microscopic asexual fungi associated with the plant substrates which make up bird nests in a fragment of the semi-deciduous seasonal forest in the “Serra na Maravilha”, Senhor do Bonfim, Bahia, Brazil.

Materials and Methods

The “Serra de Jacobina”, located in the Meridional Sertão Depression Eco-Region (Velloso

et al. 2002), is formed by a mountainous complex that extends 200 km from north to south, an extension more to the northeast of the “Chapada Diamantina”, state of Bahia, Brazil. In this complex, the “Serra da Maravilha” pertains to the city of Senhor do Bonfim, with an altitude that varies from 700 m to 1000 m and vegetation typical of the Caatinga being less expressive, being associated with a Semi-deciduous Seasonal Forest (Castelletti et al. 2003).

Sampling expeditions were carried out in a fragment of the semi-deciduous seasonal forest in the “Serra da Maravilha”, from February to July 2014. In the first expedition, demarcation of abandoned nests took place randomly as they were found, and from the composition and morphology of the nests the ethnozoological identification of them was carried out by a native of the area, who indicated the common names of the birds. These common names were compared with a list of local bird-fauna to arrive at the scientific names (Table 1).

Table 1 Number of nests studied from each bird species and substrate composition.

Birds/ Common name	No. of nests collected	Composition
<i>Arremon taciturnus</i> (Hermann 1783), pectoral sparrow	1	Straw of <i>Syagrus coronata</i> (Martius) Beccari and undetermined angiosperm twigs.
<i>Cacicus cela</i> (Linnaeus 1758), yellow-rumped cacique	1	Poaceae leaves and undetermined angiosperm twigs.
<i>Leptotila verreauxi</i> Bonaparte 1855, white-tipped dove	1	Barks, twigs and leaves from undetermined angiosperms.
<i>Sporophila nigricollis</i> (Vieillot 1823), yellow-bellied seedeater	1	Leaves and fine roots from undetermined angiosperms.
<i>Taraba major</i> (Vieillot 1816), great antshrike	1	Poaceae leaves and undetermined angiosperm twigs and leaves.
<i>Turdus rufiventris</i> Vieillot 1818, rufous-bellied thrush	5	Petioles, twigs, leaves and fine roots from undetermined angiosperms and bryophytes.
Undetermined, “tuti”	1	Leaves and roots from undetermined angiosperms.

Plant samples from the abandoned nests were collected every two months and placed in sterile plastic bags. Each sample filled the volume of plant material necessary for assembling a humid chamber in Petri dishes (150 × 18mm). The samples were taken to the “Laboratório de Biologia Molecular e Fungos” the Universidade do Estado da Bahia, Departamento de Educação, Campus VII, for processing.

At the laboratory, the material went through a modified washing, drying and incubation process created by Castañeda-Ruiz (2005), which consisted of washing the material in running water for around 20 minutes, drying the materials on a paper towel at room temperature for around 20 minutes and then incubation in Petri dishes containing moistened filter paper, followed by packing the chambers in Styrofoam boxes, covered once again with moist paper towels. After incubation and processing, the asexual microscopic fungal reproduction structures were transferred to permanent slides containing PVL (polyvinyl alcohol + lactophenol). Identification was based on classic taxonomy with the help of a basic and specialized bibliography. The sample specimens were deposited in the Herbarium at the Universidade do Estado da Bahia (HUNEB – Senhor do Bonfim Collection).

Unfortunately, by the second collecting three *Turdus rufiventris* nests, one *A. taciturnus* and one undetermined (“Tuti”) had been lost, while at the third collecting there was a loss of a *S. nigricollis* nest.

Results and Discussion

From the 11 nests, 44 species of microscopic asexual fungi were identified distributed in 36 genera. The greatest richness of species was recorded in the first sampling (32 species) followed by the second sampling (21 species) and the third sampling (19 species) (Table 2). The decreasing

number of fungal species found during the sampling is probably explained by the decreasing number of nests collected at each sampling. The decreasing number of nests sampled in the collections happened mostly due to anthropic actions, through fallen trees in the study area as well as through heavy rains that ended up knocking them down.

The *Turdus rufiventris* nests recorded the greatest richness of microscopic asexual fungi, with 28 species, followed by the *Taraba major* nest with 14 species, while the *A. taciturnus* nest had only one species of fungus recorded. Such differences could be associated with the composition of the bird nests since the *Turdus rufiventris* and *Taraba major* nests showed different substrates and that of the *A. taciturnus* had only two substrates in its composition (Table 1).

Among the species found, *Actinocladium rhodosporium*, *Alternaria alternata*, *Cryptophiale kakombensis*, *Ellisembia adscendens*, *Gyothrix circinata*, *Gyothrix microsperma*, *Tetraploa aristata*, *Thozetella cristata* and *Torula herbarum* occurred throughout the study, while the other species were found sporadically.

Most of the species found in the study are known in Brazil as associated with leaf litter materials (Marques et al. 2007, 2008, Barbosa et al. 2008, Cruz & Gusmão 2009, Almeida et al. 2012), showing that the substrates that compose the nest as well as their structures, favor colonization by microscopic asexual fungi, which was expected due to the leaf litter serving as a source of material that makes up the nests.

Dendryphion cubense constitutes a new record for South America, being previously referenced in Cuba (Matsushima 1987), Thailand (Kodsueb et al. 2008) and Taiwan (Chen & Tzean 2009). *Endophragmiella valdiviana* represents a new record for Brazil, being previously cataloged in Chile (Ellis 1963), New Zealand (Hughes 1979), Japan (Matsushima 1983), China (Wu & Zhuang 2005), Spain (Silvera-Simón et al. 2009) and Mexico (Arias et al. 2010, Abarca et al. 2013) and *Uberispora heteroseptata* constitutes a new record for Bahia, being previously registered in Brazil in São Paulo (Silva & Grandi 2008) and Pernambuco (Cruz & Gusmão 2009) (Fig. 1).



Fig. 1 – General aspect of the new records of microscopic asexual fungi species associates with bird nests in a fragment of the seasonal forest in the “Serra da Maravilha”, Senhor do Bonfim, Bahia. A. *Dendryphion cubense* B. *Endophragmiella valdiviana*. C. *Uberispora heteroseptata* (Bars = 30 μ m).

Table 2 Richness of microscopic asexual fungi associated with bird nests in a fragment of the “Serra da Maravilha”, Senhor do Bonfim, Bahia, Brasil (Tr = *Turdus rufiventris*; Lv = *Leptotila verreauxi*; At = *Arremon taciturnus*; Sn = *Sporophila nigricollis*; Tm = *Taraba major*; Cc = *Cacicus cela* and In = sp. undetermined; *New record for South America; **New record for Brazil; *** New record for Bahia).

Species	Sampling 1	Sampling 2	Sampling 3	Associated nest
<i>Actinocladium rhodosporium</i> Ehrenb.	x	x	x	Tr e Lv
<i>Alternaria alternata</i> (Fr.) Keissl.	x	x	x	Tr, Lv, Sn e Tm
<i>Beltrania rhombica</i> Penz.	x	x		Tr
<i>Beltraniella portoricensis</i> (F. Stevens) Piroz. & S.D. Patil			x	Tr
<i>Brachysporiella gayana</i> Bat.		x		Lv
<i>Chalara alabamensis</i> Morgan-Jones & E.G. Ingram	x		x	Tr
<i>Circinotrichum olivaceum</i> (Speg.) Piroz.		x		Tr
<i>Cryptophiale kakombensis</i> Piroz.	x	x	x	Tr
<i>Curvularia eragrostidis</i> (Henn.) J.A. Mey.	x			Tr e Tm
* <i>Dendryphion cubense</i> Matsush.	x			Tr
<i>Dictyoachaeta triseptata</i> (Matsush.) R.F. Castañeda	x			Tr e In
<i>Ellisemia adscendens</i> (Berk.) Subram.	x	x	x	Tr e Lv
** <i>Endophragmiella valdiviana</i> (Speg.) S. Hughes			x	Cc
<i>Fusariella atrovirens</i> (Berk.) Sacc.	x			Tm
<i>Gyrothrix circinata</i> (Berk. & M.A. Curtis) S. Hughes	x	x	x	Tr, Tm e Cc
<i>Gyrothrix microsperma</i> (Höhn.) Piroz.	x	x	x	Tr
<i>Gyrothrix podosperma</i> (Corda) Rabenh.	x			Tr
<i>Gyrothrix ramosa</i> Zucconi & Onofri			x	Tm
<i>Hermatomyces sphaericus</i> (Sacc.) S. Hughes	x	x		Lv
<i>Junewangia globulosa</i> (Tóth) W.A. Baker & Morgan-Jones	x			Lv
<i>Memnoniella subsimplex</i> (Cooke) Deighton	x			In
<i>Menisporopsis theobromae</i> S. Hughes	x		x	Cc e In
<i>Periconia byssoides</i> Pers.	x		x	Tr, Lv, Sn, Tm e Cc
<i>Periconia macrospinosa</i> Lefebvre & Aar.G. Johnson	x			Sn
<i>Periconia minutissima</i> Corda	x			Tr, Tm e In
<i>Phaeoisaria infrafertilis</i> B. Sutton & Hodges		x		Tr
<i>Pithomyces chartarum</i> (Berk. & M.A. Curtis) M.B. Ellis	x			Tr, Lv, Sn, Tm e In
<i>Pithomyces maydicus</i> (Sacc.) M.B. Ellis	x	x		Tr e Sn
<i>Pseudoacrodictys viridescens</i> (B. Sutton & Alcorn) W.A. Baker & Morgan-Jones		x		Tr
<i>Satchmopsis brasiliensis</i> B. Sutton & Hodges	x	x		Tr
<i>Spegazzinia deightonii</i> (S. Hughes) Subram.		x		Tm
<i>Spegazzinia tessarthra</i> (Berk. & M.A. Curtis) Sacc.	x	x		Sn e Tm
<i>Sporendocladia foliicola</i> (P.M. Kirk) M.J. Wingf.			x	Lv
<i>Sporidesmiella aspera</i> Kuthub. & Nawawi		x		Tr
<i>Sporidesmiella hyalosperma</i> (Corda) P.M. Kirk	x	x		Tr e Cc
<i>Stachybotrys nephrospora</i> Hansf.			x	Tm
<i>Tetraploa aristata</i> Berk. & Broome	x	x	x	Lv e Tm
<i>Thozetella cristata</i> Piroz. & Hodges	x	x	x	Tr e Lv
<i>Torula herbarum</i> (Pers.) Link	x	x	x	Tr, Sn, Tm e Cc
*** <i>Uberispora heteroseptata</i> R.F. Castañeda, Guarro & Cano	x			Tr
<i>Umbellidium radulans</i> B. Sutton & Hodges			x	Tr
<i>Veronaea coprophila</i> (Subram. & Lodha) M.B. Ellis	x		x	Tr e Tm
<i>Volutella minima</i> Höhn.	x			In
<i>Zygosporium minus</i> S. Hughes	x			At

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